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UDC 547.972

The glycosidation of flavones at the 5-OH group is rarely found in nature [1, 2]. The finding in <u>Equisetum arvense</u> L. of luteolin 5-glucoside has been reported previously [5], but later the question of the finding of this compound was placed under doubt [1, 4].

Continuing a study of the flavonoids of the epigeal part of E. arvense L. (field horsetail) we have isolated from an ethyl acetate extract by chromatography on a polyamide sorbent a white crystalline substance with mp 255-256°C, [ $\alpha$ ]<sup>20</sup>-22.7° (c 0.175; methanol)  $\lambda_{\text{max}}^{\text{CH}_3\text{OH}}$  262, 333 (log  $\epsilon$ , 413; 416);  $\lambda_{\text{max}}^{\text{(+AlCl}_3)}$  262, 335;  $\lambda_{\text{max}}^{\text{(+AlCl}_3+HCl)}$  263,335;  $\lambda_{\text{max}}^{\text{(+CH}_3\text{COONa)}}$  271, 305, 370;  $\lambda_{\text{max}}^{\text{(+CH}_3\text{ONa)}}$  270, 330, 390 nm. The band of the valence vibrations of a carbonyl group is present at 1630 cm<sup>-1</sup> and that of hydroxy groups in the 3300-3400-cm<sup>-1</sup> region.

From the results of quantitative acid and enzymatic hydrolytic cleavage it can be seen that the substance under investigation is a monoside containing D-glucose as the carbohydrate component and apigenin as the aglycone [5].

The compound under consideration possesses a bright-blue fluorescence in UV light, which is characteristic for flavone 5-glycosides [1]. The absence of a bathochromic shift in the presence of AlCl<sub>3</sub> for the glycoside, and also the absence of the signal of the proton of the 5-OH group in the weak field of the PMR spectrum (with dimethyl sulfoxide as solvent, B 487B radiospectrometer) in comparison with apigenin shows that the glucose is attached at position 5.

The capacity of the glycoside for being split by  $\beta$ -glucosidase shows the  $\beta$ -configuration of the glycosidic bond. On comparing the molecular rotations of the glycoside with the corresponding figures for phenyl glycosides it can be seen that the carbohydrate substituent is  $\beta$ -D-glucopyranose. The pyranose form is also confirmed by the results of IR spectroscopy (1100, 1070, and 1030 cm<sup>-1</sup>) [16]. A doublet at  $\delta$  5.02 ppm (1H, J=6 Hz) corresponds to the anomeric proton of the  $\beta$ -glucose in position 5 [7].

Thus, the compound under investigation has the structure of apigenin 5-O- $\beta$ -D-glucopyranoside.

Apigenin 5-glucoside has been isolated previously from Amorpha fruticosa [8] and was later synthesized, but its physicochemical constants, with the exception of the melting point, were not given. The melting point of the apigenin 5-glucoside that we have identified (255-256°C) differs from that given in the literature (295°C) [8, 9].

This is the first time that apigenin 5-O- $\beta$ -D-glucopyranoside has been found in the family Equisetaseae.

## LITERATURE CITED

- 1. J. B. Harborn, Phytochemistry, 6, 1569 (1967).
- 2. V. A. Bandyukova and É. T. Avanesov, Khim. Prirodn. Soedin., 413 (1972).
- 3. H. Nakamure and G. Hukuti, J. Pharm. Soc. Jap., 60, 449 (1940).
- 4. N. A. M. Salleh, W. Majak, and G. H. N. Towers, Phytochemistry, 11, 1095 (1973).

Irkutsk Institute of Organic Chemistry, Siberian Branch, Academy of Sciences of the USSR. Translated from Khimiya Prirodnykh Soedinenii, No. 5, pp. 666-667, September-October, 1974. Original article submitted April 30, 1974.

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- 5. A. I. Syrchina, M. G. Voronkov, and N. A. Tyukavkina, Khim. Prirodn. Soedin., 671 (1973)
- 6. I. L. Kovalev and V. I. Litvinenko, Khim. Prirodn. Soedin., 233 (1965).
- 7. T. J. Mabry, K. R. Markham, and M. B. Thomas, The Systematic Identification of Flavonoids, Springer, New York (1970), p. 269.
- 8. R. Goto and M. Taki, J. Pharm. Soc. Jap., <u>58</u>, 933 (1938).
- 9. G. Zemplen and L. Mester, Ber., 76, 776 (1943).